

The Effect of Different Concentrations of Some Nanoparticles and Seaweed Extract on the Vegetative Growth of Rosmary [*Salvia Rosmarinus L.*] Plant

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Abstract:

The presented study to investigate the response of rosmarinus (Salvia rosmarinus L.) plants on the influence of nanoparticles (zinc and magnesium) and seaweed extract carried out in 2024 on fields in wasit university, College of Sciences, Iraq. The treatment construction was a perfectly randomized factorial experiment (5×5×3) in completely randomized design, done with three replications. The rosemary plants treated with nanoparticles (zinc oxide) with concentrations (0, 0.03, 0.05) mg/ml and (magnesium oxide) with concentrations (0, 0.02, 0.04) mg/ml and rosmarinus treated with seaweed extract with concentrations (0, 50, 100, 150, 200) gm/L. Studying plant height (cm), total chlorophyll (μmol m⁻²), percentage of dry matter (%), leaf surface area (cm².plant⁻¹), nitrogen (%), phosphorus (%) and potassium percentage. Based on results, adding nano zinc and nano magnesium and seaweed extract to the rosemary plant significantly increased all traits, regardless of vegetable characteristics or chemical content achieved with highest concentrations using highest effects whether alone or with two interactions.

Keywords: Rosemary, Nanoparticles, Seaweed extract, Nano zinc, Nano magnesium

Key findings: In the rosemary plants, adding nanoparticles and seaweed extract and their interactions highly improved all vegetative and biochemical traits

1.Introduction

Rosemary (*Rosmarinus officinalis* L.) is an evergreen brush belonging to the Lamiaceae family. In natural conditions, it can reach from approx. 1 m to even 2.5 m in height. The stems are quadrangular, erect, and tend to lignify in the 2nd year. They are densely covered with small needle-like leaves without stalks (sessile); the leaves are linear with entire slightly revolute margins that are dark-green above, lighter and tomentose beneath. Rosemary blooms from early June to August. The flowers are very small, white or purplish-blue, gathered in terminal racemose inflorescences (Pawłowska K et al., 2020). Rosemary is cultivated in the Mediterranean region, as well as in the former Yugoslavia, along the Black Sea coast, in the USA and in Mexico (Pawłowska K et al., 2020). It is best planted in a sunny position and must be protected from cold winds (mainly in temperate climate zones). The plant reproduces predominantly sexually. The soil needs to be loose and loamy with excellent drainage. When grown in such conditions, the plant will have a more intense aroma, though at the expense of size. Leaves produced at the beginning of bloom contain the highest amounts of oil, and is when they should be collected. Apart from the leaves, non-woody sprigs are also used. After harvesting, the material is air dried, in the shade preferably, at 30–35°C (Pawłowska K et al., 2020). The use of rosemary spice in food increases the secretion of gastric juice, improves digestion. For this importance of rosemary there has been a trend towards using modern technologies in treating rosemary, including nanotechnology, which is considered one of the modern technologies used to treat the plant due to its effective and positive impact in increasing the productivity and effectiveness of this plant.

The prefix 'nano' is referred to a Greek prefix meaning 'dwarf' or something very small and depicts one thousand millionth of a meter (10⁻⁹ m). We should distinguish between nanoscience, and nanotechnology. (Gnani A et al., 2015).

Assessing the role of nano-fertilizers and elicitor as a strategy is important for reducing the negative effects of drought stress on plants. The physicochemical properties, such as outstanding refractive index, excellent corrosion resistance, high thermal conductivity, low electrical conductivity, physical strength, stability, flame resistance, dielectric resistance,

mechanical strength and excellent optical transparency, magnesium oxide (MgO) is an eco-friendly, economically feasible and industrially important nanoparticle(Abinaya S et al ., 2021).It is regarded as a promising high-surface-area heterogeneous catalyst support, additive, and promoter for a variety of chemical reactions due to its unique properties, which include stoichiometry and composition, cation valence and redox properties, acid-base character and crystal and electronic structure , (Julkapli N M & Bagheri S , 2016) .

Nano magnesium -fertilizers include the nutrients with the size 30 to 40 nanometer and can carry nutrients properly due to their high surface.The use of nano-fertilizers to control the release of nutrients accurately can be considered as an effective step toward achieving sustainable and eco-friendly agriculture. Further, Mg is an important task for plants based on quantitative and qualitative, energy conservation, protein synthesis, structure stability, and sugar accumulation, as well as controlling pH in the cell and a cofactor for many enzymes (Blasco et al ., 2015) .Zinc is required for chlorophyll production, pollen function, fertilization, and germination(Kaya and Higgs 2002; Pandey, Pathak, and Sharma 2006; Cakmak 2008). It plays an important role in biomass production (Kaya and Higgs 2002). Among the micronutrients, Zn and Mn can affect the susceptibility of plants to drought stress (Khan, McDonald, and Rengel 2003). A number of mechanisms may underlie Zn efficiency (Rengel 2001).

Seaweed extracts contain trace elements and plant growth hormones IAA and IBA growth stimulators such as auxin, gibberellins and cytokinin required by plants. It is also reported that seaweed extracts are rich in vitamins, amino acids, trace elements (Fe, Cu, Co, Ni, Zn and Mn) (Booth E ,1969). Seaweed extracts have nitrogen, phosphorus and higher amount of water soluble potash, other minerals and trace elements in a readily absorbable by plants. They control deficiency diseases. Today is being used of low concentration of seaweed extracts in combination with commercial fertilizer. It has been used more effectively in the world in promoting plant growth (Chatterji, 2004).

seaweed extracts have been highly explored for possible use in crop production for improving biomass yield and produce quality. These extracts have been shown to positively affect seed germination and plant growth at all stages up to harvest and even post-harvest (Ali O et al ., 2019 ; Ali O et al ., 2021). The improved rooting architecture could be a result of small levels of phytohormones present in the extracts such as auxins as well as various stimulatory processes engaged in the plant system upon treatment with these extracts (Crouch IJ et al ., 1992).

2.MATERIALS AND METHODS:

The experimentation commenced in 15/11/2023 at the private archard field in Wasit University ,college of science The soil mixture was placed in the pot, the capacity of each anvil (10kg) placed in the pots and watered with water and on foliar spraying operations were carried out and soil spraying according to the previously mentioned concentrations Spraying plant leaves until the entire leaf is covered with concentrations of nanocomposites and seaweed extract, and repeated spraying and addition after a month of the first treatment agricultural service operations of plant watering and soil fertilization were carried out throughout the duration of the experiment . In this study, nanocomposites were prepared and prepared at certain concentrations depending on the manufacturer concentrations (0,0.02,0.03,0.04,0.05) mg/ml by dissolving the required concentrations with 5 ml of ethanol . The volume is supplemented to 500ml of deionized water to obtain the above required concentrations , as well as the use of seaweed extract in concentrations. (0,50 , 100 , 150 , 200) gm/ L and attended concentrates dissolved with distilled water and complete the volume to 1 liter.

There were 75 panels the foliar spraying of nanoparticles at concentrations (0,0.02 ,0.04 ,0.03 , 0.05) mg/ml and seaweed extract at concentrations (0,50 , 100 , 150 , 200) gm/L .Foliar spraying at one time ensure for treatment in the vegetative stage of the plant After two weeks Foliar spraying was done for the second time and after two week we begin the methods , estimation of plant height , chlorophyll(Minotti P et al ., 1994) , percentage of dry matter (Teye E et al ., 2011) , leaf surface area(Lang, A R G ,1973)., nitrogen percentage(Gresser M S & Parsons J M ,1979), and phosphorus percentage((Pratt P F & Chapman H D ,1961 and potassium percentage (A.O.A.C ,1990).

2.1 STATISTICAL ANALYSIS:

The Randomized complete block design (RCBD) analysis of variance test helped examin how statistically different treatments differed. The the least significant differences (LSD) at 0.05 % levels assessed mean comparison (Steel and Torrie , 1980).

3.RESULTS AND DISCUSSION:

3.1 Average Plant Height

Table 1 shows that nano compound significantly affects average plant height. When the concentration of nano zinc 0.05 mg/ml the mean plant height 23.33 cm. Plants treated with control grew the fastest 26.33 cm. Nano magnesium also significantly affected average plant height. Similarly, when the concentration of nano magnesium 0.02, it increased the average plant height. The greatest average of plant height was created by Seaweed extract at 150 gm /L (6.811)cm, compared to 50 (25.07) cm and the control (23.93) cm. Nanocompound and Seaweed extract had a significant interaction effect (see Table 1). Plants treated with 0.03 mg/ml of nanozinc and 50 of Seaweed extract average of plant height.

Table (1) : Effect of some Nano compounds (Nanomagnesium , Nanozinc) and Seaweed extract and the interactions between them on average plant height (cm)

Seaweed Extract (E)	Nanocompounds					Average of Seaweed Extract (E)
	0	0.02(MgO) mg/m l	0.04 (Mg O)m g/l	0.03 (ZnO) mg/ml	0.05 (ZnO) mg/m l	
0	19.67	26.00	28.67	29.67	27.67	23.93
50gm/L	26.00	25.67	22.33	32.00	23.67	25.07
100gm/L	27.33	26.67	26.67	22.00	19.00	25.60
150gm/L	25.33	24.67	24.33	22.67	26.67	26.07
200gm/L	21.33	22.33	26.00	24.00	23.00	24.00
Average of Nanocompounds (N)	26.33	25.93	24.73	24.33	23.33	Nanocompounds (N)
LSD (P=0.05)	3.217					LSD (P=0.05)
						3.217

3.2 Estimation of total chlorophyll ($\mu\text{mol m}^{-2}$)

Table 2 shows that nano compound significantly affects average total chlorophyll . When the concentration of nano zinc 0.05 mg/ml the mean total chlorophyll (1.161) $\mu\text{mol m}^{-2}$. Plants treated with control grew the fastest (1.153) $\mu\text{mol m}^{-2}$ Nano magnesium also significantly affected average total chlorophyll . Similarly, when the concentration of nano magnesium 0.02 mg/ml , it increased the average total chlorophyll. The greatest average of total chlorophyll was created by Seaweed extract at 150 gm /L (1.245) compared to 50 (1.165) $\mu\text{mol m}^{-2}$ and the control (1.231) $\mu\text{mol m}^{-2}$ Nanocompound and Seaweed extract had a significant interaction effect (see table 3) . Plants treated with 0.03 mg/ml of nanozinc and 50 mg/ml of Seaweed extract average of total chlorophyll.

Table (2) : Effect of some of nanocompounds and seaweed extract and the interactions between them on average Total chlorophyll (mg/m^3)

Seaweed Extract (E)	Nanocompounds`					Average of Seaweed Extract (E)
	0	0.02(MgO) mg/ml	0.04 (MgO) mg/ml	0.03 (ZnO) mg/ml	0.05 (ZnO) mg/ml	
0	1.063	1.360	1.273	1.307	1.150	1.231
50 gm/L	1.140	1.120	1.093	1.287	1.187	1.165
100gm/L	1.130	1.180	1.327	1.313	1.227	1.235
150gm/L	1.313	1.347	1.233	1.200	1.133	1.245
200gm/L	1.120	1.240	1.157	1.233	1.107	1.171

Average of Nanocompounds (N)	1.153	1.249	1.268	1.217	1.161	Nanocompounds (N)
LSD (P=0.05)	0.0576					LSD (P=0.05)
						0.0576
Two-way interaction LSD (P=0.05)	0.1288					

3.3 The percentage of dry matter of vegetative parts of the plant (%)

Table 3 shows that nano compound significantly affects average the percentage of dry matter of the vegetative parts of the plant (%). When the concentration of nano zinc 0.05 mg/ml the mean the percentage of dry matter of the vegetative parts of the plant 88.9% . Plants treated with control grew the fastest 81.4 % Nano magnesium also significantly affected average the percentage of dry matter of the vegetative parts of the plant %. Similarly, when the concentration of nano magnesium 0.02 mg/ml, it increased the average the percentage of dry matter of the vegetative parts of the plant (%). The greatest average of the percentage of dry matter of the vegetative parts of the plant (%) was created by Seaweed extract at 150 gm /L (105.8%) , compared to 50gm/L (73.7%) and the control (105.8%) . Nanocompound and Seaweed extract had a significant interaction effect (see table7). Plants treated with 0.03 mg/ml of nanozinc and 50 of Seaweed extract average of the percentage of dry matter of the vegetative parts of the plant (%).

Table (3) :Effect of some of Nano compounds (Nanomagnesium , Nanozinc) and seaweed extract and the interactions between them on average The percentage of dry matter of the vegetative parts of the plant (%)

Seaweed Extract (E)		Nanocompounds				Average of Seaweed Extract (E)
	0	0.02(MgO) mg/ml	0.04 (MgO) mg/ml	0.04 (ZnO) mg/ml	0.05 (ZnO) mg/ml	
0	43.3	104.6	42.1	50.2	139.3	75.9
50 gm/L	85.6	75.6	79.9	108.6	18.6	73.7
100gm/L	79.9	61.3	66.1	147.5	70.7	85.1
150gm/L	104.2	80.3	72.1	126.4	146.1	105.8
200gm/L	104.2	79.9	154.5	87.8	70.0	99.3
Average of Nanocompounds (N)	81.4	82.3	104.1	82.9	88.9	Nanocompounds (N)
LSD (P=0.05)		47.70				LSD (P=0.05)
						47.70
Two-way interaction LSD (P=0.05)		106.67				

3.4 Leaf surface area

Table 4 shows that nano compound significantly affects average leaf surface area . When the concentration of nano zinc 0.05 mg/ml the mean leaf surface area(78.9) cm².plant-1 . Plants treated with control grew the fastest (73.5) cm².plant-1 . Nano magnesium also significantly affected average leaf surface area . Similarly, when the concentration of nano magnesium 0.02 mg/ml, it increased the average leaf surface area. The greatest average of leaf surface area was created by

Seaweed extract at 150 gm /L (74.3) cm².plant-1 compared to 50gm/L (73.9) cm².plant-1 and the control (72.7) cm².plant-1. Nanocompound and Seaweed extract had a significant interaction effect (see Table 4).Plants treated with 0.03 mg/ml of nanozinc and 50gm/L of Seaweed extract averave of leaf surface area.

Table (4) :Effect of some of nanocompunds (Nanomagnesium , Nanozinc) and seaweed extract and the interactions between them on average Leaf surface area (cm².plant-1)

Seaweed Extract (E)	Nanocmpounds					Average of Seaweed Extract (E)
	0	0.02(MgO) mg/ml	0.04 (MgO) mg/ml	0.03 (ZnO) mg/ml	0.05 (ZnO) mg/ml	
0	88.4	76.0	65.4	64.7	69.0	72.7
50gm/L	71.1	63.7	65.0	49.1	70.7	73.9
100gm/L	72.3	55.0	61.0	90.7	91.3	74.1
150gm/L	67.2	78.3	52.0	85.3	88.7	74.3
200gm/L	68.4	81.3	65.0	68.0	74.7	71.5
Average of Nanocompounds (N)	73.5	70.9	81.6	61.7	78.9	Nanocompounds (N)
LSD (P=0.05)	13.14					LSD (P=0.05)
						13.14
Two-way interaction LSD (P=0.05)	29.38					

3.5Nitrogen (%)

Table 4 shows that nano compound significantly affects average nitrogen content (%). When the concentration of nano zinc 0.05 mg/ml the mean nitrogen (%) 1.043 %. Plants treated with control grew the fastest 1.040 (%). Nano magnesium also significantly affected average nitrogen content (%). Similarly, when the concentration of nano magnesium 0.02 mg/ml, it increased the average nitrogen (%). The greatest average of plant height was created by Seaweed extract at 150 gm /L (0.912) % compared to 50 gm/L (1.064)% and the control (0.987)%. Nanocompound and Seaweed extract had a significant interaction effect (see Table(4).Plants treated with 0.03 mg/ml of nanozinc and 50 gm/L of Seaweed extract averave of nitrogen (%).

Table (5) :Effect of some of Nano compounds (Nanomagnesium , Nanozinc) and seaweed extract and the interactions between them on Nitrogen (%)

Seaweed Extract (E)	Nanocmpounds					Average of Seaweed Extract (E)
	0	0.02(MgO) mg/ml	0.04 (MgO) mg/ml	0.03 (ZnO) mg/ml	0.05 (ZnO) mg/ml	
0	1.097	1.213	0.910	0.700	1.013	0.987
50gm/L	1.073	1.307	1.050	0.957	0.933	1.064
100gm/L	1.120	0.700	1.013	1.120	1.587	1.108
150gm/L	1.013	0.910	0.863	0.910	0.863	0.912
200gm/L	0.897	1.260	1.633	1.073	0.817	1.136
Average of Nanocompounds (N)	1.040	1.078	0.952	1.094	1.043	Nanocompounds (N)

LSD (P=0.05)	0.2427	LSD (P=0.05)
		0.2427
Two-way interaction LSD (P=0.05)	0.5426	

3.6 Phosphorus (%)

Table 6 shows that nano compound significantly affects average Phosphorus (%). When the concentration of nano zinc 0.05 mg/ml the mean Phosphorus (0.2689 %). Plants treated with control grew the fastest (0.2391%) . Nano magnesium also significantly affected average Phosphorus (%). Similarly, when the concentration of nano magnesium 0.02 mg/ml, it increased the average phosphorus (%). The greatest average of Phosphorus (%) was created by Seaweed extract at 150 gm /L (0.2552%), compared to 50gm/L (0.2535%) and the control (0.2589%). Nanocompound and Seaweed extract had a significant interaction effect (see Table 6). Plants treated with 0.03 mg/ml of nanozinc and 50 of Seaweed extract average of Phosphorus (%) .

Table (6) :Effect of some of Nano compounds (Nanomagnesium , Nanozinc) and seaweed extract and the interactions between them on average Phosphorus percentage (%)

Seaweed Extract (E)	Nanocompounds					Average of Seaweed Extract (E)
	0	0.02(MgO) mg/ml	0.04 (MgO) mg/ml	0.03 (ZnO) mg/ml	0.05 (ZnO) mg/ml	
0	0.2133	0.2347	0.2430	0.3453	0.2583	0.2589
50gm/L	0.2643	0.2897	0.2663	0.2583	0.1890	0.2535
100gm/L	0.2650	0.3580	0.1277	0.2183	0.2960	0.2530
150gm/L	0.2577	0.2283	0.2153	0.2480	0.3267	0.2552
200gm/L	0.1953	0.2563	0.2947	0.2720	0.2747	0.2586
Average of Nanocompounds (N)	0.2391	0.2734	0.2684	0.2294	0.2689	Nanocompounds (N)
LSD (P=0.05)	0.05102					LSD (P=0.05)
						0.05102
Two-way interaction LSD (P=0.05)	0.11407					

3.7 Potassium (%)

Table 7 shows that nano compound significantly affects average Potassium (%) .

Table (7) :Effect of some of Nanocompounds (Nanomagnesium , Nanozinc) and seaweed extract and the interactions between them on average Potassium percentage (%)

Seaweed Extract (E)	Nanocompounds					Average of Seaweed Extract (E)
	0	0.02(MgO) mg/ml	0.04 (MgO) mg/ml	0.03 (ZnO) mg/ml	0.05 (ZnO) mg/ml	
0	0.896	0.536	1.406	1.011	1.092	0.988
50gm/L	0.873	0.906	1.037	1.182	0.772	0.954
100gm/L	1.493	1.058	0.890	0.657	0.826	0.985
150gm/L	0.959	1.015	0.766	1.367	0.912	1.004
200gm/L	0.811	0.708	0.840	0.932	0.963	0.851
Average of Nanocompounds	1.006	0.845	1.030	0.988	0.913	Nanocompounds

(N)						(N)
LSD (P=0.05)	0.3033					LSD (P=0.05)
						0.3033
Two-way interaction LSD (P=0.05)	0.6782					

When the concentration of nano zinc 0.05 mg/ml the mean Potassium (0.913 %) . Plants treated with control grew the fastest (0.913%). Nano magnesium also significantly affected Potassium (%). Similarly, when the concentration of nano magnesium 0.02 mg/ml, it increased the average Potassium (%). The greatest average of Potassium (%)was created by Seaweed extract at 150 gm /L (1.004%), compared to 50 (0.954%) and the control (0.988%).Nanocompound and Seaweed extract had a significant interaction effect (see Table 7).Plants treated with 0.03 mg/ml of nanozinc and 50 of Seaweed extract average of Potassium (%).

Comparison of nano-compounds and seaweed extract on the quantitative yield of plants with other reports from different parts of the world represents the difference characteristics in this species is in all Generally, vegetative growth such as wet and dry weight of leaves, stems and whole plant.

when the plant treated with nanoscale ZnO particles that increase in plant height can be ascribed to higher precursor activity of nanoscale zinc in auxin production (Kobayashi and Mizutani 1970). It was found that the nanocomposites containing zinc enhanced the growth indicators of the plant, which is consistent with the results of the current study, which showed that the foliar application of nano zinc Khoshgoftarmanesh et al. (2020) led to a significant increase in plant height, leaf area, and dry weight, compared to control plants. Zinc nanocomposites can improve the absorption and assimilation of nutrients in plants, which is consistent with the results of the current study that foliar application of zinc nanocomposites) Fakheri et al ., 2020) . It resulted in a significant increase in zinc concentration in plant leaves compared to untreated plants. Moreover, nano zinc compounds can enhance the availability and uptake of other essential nutrients, including nitrogen, phosphorus and potassium, which leads to improved nutrient status and overall plant growth according to (Seyedbagheri et al., 2021).

On the other hand, foliar spraying of nanocomposites directly on the leaves, this method provides effective pathways for delivering nutrients to plants, and foliar spraying allows rapid absorption of nutrients through the leaf surface, while fertilization ensures uniform distribution of nutrients to the root zone. The significant superiority obtained from nanocomposites on rosemary plants was consistent with what was obtained by a study on rosemary plants on the effect of foliar addition of nano zinc compounds (Borzouei et al., 2017) on rosemary plants, which found that the use of nano zinc led to a significant increase in plant height, leaf area, fresh ,dry weights, and improved chlorophyll pigments compared to the control group.

Regarding with results nitrogen and phosphorus concentration the highest concentrations of nitrogen, phosphorus and potassium were recorded in the leaves of plants treated with nanocomposites compared to the comparison group, as the study agreed with (Baghbani et al., 2018) .The interpretation of the moral superiority of foliar spraying with nano compounds in the current study was consistent with the study of (Soltani , 2017) on rosemary plants, as foliar spraying using nano zinc led to a significant increase in plant height and leaf area compared to the comparison group, in addition, the addition of nano zinc improved the chlorophyll content, photosynthesis rate, and accumulation of essential nutrients such as nitrogen, phosphorus, and potassium.

The increases in leaf surface area, dry weight and plant height concord with growth-stimulating effects of SWE reported in several studies on other horticultural crops (Kumari et al., 2011; Alam et al., 2013; Mattner et al., 2013; Hernández-Herrera et al., 2014). . Seaweed extract treatment enhanced both root :shoot Although they may contain different levels of minerals bio stimulants are unable to provide all the nutrients needed by a plant in required quantities (Schmidt and others 2003);however, their main benefit is to improve plant mineral uptake by the roots (Vernieri and others 2005) and in the leaves (Mancuso and others 2006). Both treatments resulted in higher chlorophyll levels versus controls measured on both days. The results imply that the enhanced leaf chlorophyll content of plants treated with seaweed extract might depend on betaines present in the extract (Blunden and others 1997).

(Blunden et al .,1996) found that the application of seaweed extract had a higher chlorophyll content than unapplied treatments, which might be attributed to the reduction in chlorophyll degradation caused by betaines in the seaweed extract.

Besides, some researchers found that the application of seaweed increased the leaf surface area, (Lola-Luz, T et al., 2013)

The effect of seaweeds on plant growth and height (Kardiner *et al.*, 1969). This result was consistent with what Sadeghi et al. (2009) in basil plants.. The positive effect of the extract of seaweed kelp on the studied traits may be due to its hormonal nature, as it contains auxins and cytokinins, which leads to an increase in the concentration of these hormones inside the plant, which is positively reflected on vegetative growth (Crouch et al., 1992). It also stimulates the growth of the root system, which helps in increasing the absorption of nutrients from the soil solution (Beckett and Van-Stden, 1989), which is positively reflected In increasing the vegetative growth indicators, and this is consistent with what Almohammed et al. (2014) reached on the black seed plant.

We also find that the use of seaweed extracts leads to a significant increase in plant height, leaf area and chlorophyll content in the leaves, according to (Reitz, S.R. & J. T. Trumble, 1996) in the bean plant, (Reitz, S.R. & J. T. Trumble, 1996) in the cucumber plant and (Potter G, 2005) in the lettuce plant when using other seaweed extracts. This increase in the vegetative growth characteristics resulting from treatment with seaweed extracts may be due to the fact that these extracts contain auxins, amino acids and other plant hormones, which stimulate the growth and development of the vegetative and root groups, increase the resistance of plants to stress and prevent the oxidation of vitamin C and E Which is found in chloroplasts, which increases the efficiency of the photosynthesis process (O'Dell C, 2003), as mentioned (Potter G, 2005) These extracts work to form a branched root system and thus stimulate the process of absorbing water and nutrients better from the soil and form a good root system. As mentioned (Reeta KA & AK Bhatnager, 2011) these extracts contain iron, magnesium and nitrogen, which are part of the chlorophyll molecule, which helps in the photosynthesis process. This increase in the photosynthesis process gave good vegetative growth.

These results are in harmony with those mentioned by Gharib et al. (2014) on rosemary they showed that foliar application of seaweed on rosemary plant was positively active on microelement content and photosynthetic pigments (chlorophyll) compared to the untreated plant (Pise and Sabale, 2010) showed that carbohydrate, and proteins, free amino acids, polyphenols and nitrogen content increased in seaweed treated plants. In the same direction, there was significant effect due to seaweed extract on chlorophyll, carbohydrates, nitrogen, phosphorous and potassium in leaves of artichoke plants (Saif Eldeen, 2014).

4.CONCLUSION:

According to the results of the previously mentioned study, The effect of different concentrations of some nanoparticles and seaweed extract on the vegetative growth of rosemary [*Salvia rosmarinus* L.] plant to the plant significantly increased all traits, regardless of whether the characteristics of vegetable or chemical activity achieved the highest concentrations used the highest effects, whether alone or in combination with two other compounds..

5:COMPETING INTERESTS

Authors have declared that no competing interests exist.

References:

1. A.O.A.C. (1990). Official Methods of Analysis. 15th Edition; Association of Official Analytical Chemist; Washington DC.
2. Abbas, R. J. (2010). Effect of using fenugreek, parsley and sweet basil seeds as feed additives on the performance of broiler chickens. Int. J. Poult. Sci, 9(3), 278-282.
3. Abinaya, S., Kavitha, H. P., Prakash, M., & Muthukrishnaraj, A. J. S. C. (2021). Green synthesis of magnesium oxide nanoparticles and its applications: A review. Sustainable Chemistry and Pharmacy, 19, 100368.
4. Alam, M.Z., Braun, G., Norrie, J., Hodges, D.M., 2013. Effect of *Ascophyllum* extract application on plant growth, fruit yield and soil microbial communities of strawberry. Can. J. Plant Sci. 93, 23–36.
5. Ali, O., Ramsubhag, A., & Jayaraman, J. (2019). Biostimulatory activities of *Ascophyllum nodosum* extract in tomato and sweet pepper crops in a tropical environment. PLoS One, 14(5), e0216710.
6. Ali, O., Ramsubhag, A., & Jayaraman, J. (2021). Biostimulant properties of seaweed extracts in plants: Implications towards sustainable crop production. Plants, 10(3), 531.

7. ALmohammed, A. N.; A. F. Almehemdi and R. K. Al ajeelee (2014). Impact of Bat Guano *Otonycteris hemprichii* Camd and Seaweed Extract on Some Growth and Yield Traits of Barakaseed *Nigella Sativa* L. Journal of Biology, Agriculture and Healthcare, 4(1):565.
8. Baghbani, R., Heidari, M. and Pirdashti, H. (2018). The effect of foliar application of nano-zinc and nano-manganese on morpho-physiological characteristics and essential oil composition of rosemary (*Rosmarinus officinalis* L.). Journal of Essential Oil Bearing Plants, 21(6), 1703-1712
9. Beckett, R. P. and J. van- Staden (1989). The effect of seaweed concentrate on the growth and yield of potassium stressed wheat. Plant and Soil 116: 29 -36.
10. Blasco-Algora, S., Masegosa-Ataz, J., Gutiérrez-García, M. L., Alonso-López, S., & Fernández-Rodríguez, C. M. (2015). Acute-on-chronic liver failure: Pathogenesis, prognostic factors and management. World journal of gastroenterology, 21(42), 12125.
11. Blunden, G., Jenkins, T and Liu, Y.W. (1997). Enhanced leaf chlorophyll levels in plants treated with seaweed extract. J. Appl. Phycol. 8, 535–543.
12. Booth, E. (1969). The manufacture and properties of liquid seaweed extracts. In Proc Int Seaweed Symp (Vol. 6, pp. 655-662).
13. Borzouei, A., Kafi, M., Khoshgoftarmanesh, A. H., Schulin, R. and Afyuni, M. (2017). Effect of foliar application of nano-sized zinc and manganese chelates on the growth and physiological traits of rosemary (*Rosmarinus officinalis* L.). Scientia Horticulturae, 226, 23-30.
14. Cakmak, I. (2008). Enrichment of cereal grains with zinc: agronomic or genetic biofortification?. Plant and soil, 302, 1-17.
15. Chatterji, A., Dhargalkar, V. K., Sreekumar, P. K., Parameswaran, P. S., Rodrigues, R., & Kotnala, S. (2004). Anti-influenza activity in the Indian Seaweeds-A preliminary investigation.
16. Fakheri, B., Askari, M. and Galavi, M. (2020). Effects of zinc and manganese nanoparticles on physiological traits, essential oil yield and composition of *Satureja hortensis* L. Industrial Crops and Products, 152, 112477-112490.
17. Gharib, F. A. E. L., Zeid, I. M., Salem, O. M. A. H., & Ahmed, E. Z. (2014). Effects of *Sargassum latifolium* extract on growth, oil content and enzymatic activities of rosemary plants under salinity stress. Life sci. J, 11(10), 933-945.
18. Gnach, A., Lipinski, T., Bednarkiewicz, A., Rybka, J., & Capobianco, J. A. (2015). Upconverting nanoparticles: assessing the toxicity. *Chemical Society Reviews*, 44(6), 1561-1584.
19. Gresser, M. S., & Parsons, J. M. (1979). Sulfuric-perchloric acid digestion of some nutrients. Egyptian Journal of Horticulture, 25, 55-70.
20. Hernández-Herrera, R.M., Santacruz-Ruvalcaba, F., Ruiz-López, M.A., Norrie, J., Hernández-Carmona, G., 2014. Effect of liquid seaweed extracts on growth of tomato seedlings (*Solanum lycopersicum* L.). J. Appl. Phycol. 26, 619–628.
21. John, M. K. 1971. The distribution of inorganic phosphorus in some soils of Western Canada and its relationship to soil characteristics; Soil Science and Plant Nutrition. 17(1): 9-14.
22. Julkapli, N. M., & Bagheri, S. (2016). Developments in nano-additives for paper industry. Journal of wood science, 62, 117-130.
23. Kardiner A., L'individu dans sa société. Essai d'anthropologie psychanalytique, Gallimard, Paris, 1969 [1939].
24. Kaya, C., & Higgs, D. (2002). Response of tomato (*Lycopersicon esculentum* L.) cultivars to foliar application of zinc when grown in sand culture at low zinc. Scientia Horticulturae, 93(1), 53-64.
25. Khan, H. R., McDonald, G. K., & Rengel, Z. (2003). Zn fertilization improves water use efficiency, grain yield and seed Zn content in chickpea. Plant and Soil, 249, 389-400.
26. Khoshgoftarmanesh, A. H., Khoshgoftarmanesh, Z. A., Karami, M. R. and Hajhashemi, S. (2020). Effect of zinc and manganese nanoparticles on the growth, chlorophyll content, and macro and micronutrient accumulation of rosemary (*Rosmarinus officinalis* L.). Communications in Soil Science and Plant Analysis, 51(6), 782-790.
27. Kobayashi, Y., & Mizutani, S. (1970). Studies on the Wilting Treatment of Corn Plants: III. The influence of the artificial auxin control in nodes on the behavior of rooting. Japanese Journal of Crop Science, 39(2), 213-220.
28. Kumari, R., Kaur, I., & Bhatnagar, A. K. (2011). Effect of aqueous extract of *Sargassum johnstonii* Setchell & Gardner on growth, yield and quality of *Lycopersicon esculentum* Mill. Journal of Applied Phycology, 23, 623-633.

29. Lang, A. R. G. (1973). Leaf orientation of a cotton plant. *Agricultural Meteorology*, 11, 37-51.
30. Lola-Luz, T., Hennequart, F., & Gaffney, M. (2013). Enhancement of phenolic and flavonoid compounds in cabbage (*Brassica oleraceae*) following application of commercial seaweed extracts of the brown seaweed, (*Ascophyllum nodosum*). *Agricultural and food science*, 22(2), 288-295.
31. Longnecker, N. E., & Robson, A. D. (1993). Distribution and transport of zinc in plants. In *Zinc in Soils and Plants: Proceedings of the International Symposium on 'Zinc in Soils and Plants' held at The University of Western Australia, 27–28 September, 1993* (pp. 79-91). Dordrecht: Springer Netherlands.
32. Mancuso, S., Azzarello, E., Mugnai, S., & Briand, X. (2006). Marine bioactive substances (IPA extract) improve foliar ion uptake and water stress tolerance in potted *Vitis vinifera* plants. *Advances in Horticultural Science*, 20(2), 156-161.
33. Mattner, S.W., Wite, D., Riches, D.A., Porter, I.J., Arioli, T., 2013. The effect of kelp extract on seedling establishment of broccoli on contrasting soil types in southern Victoria. *Aust. Biol. Agric. Hort.* 29, 258–27
34. Minotti, P. L., Halseth, D. E., & Siczka, J. B. (1994). Field chlorophyll measurements to assess the nitrogen status of potato varieties. *HortScience*, 29(12), 1497-1500.
35. O'Dell, C. (2003). Natural plant hormones are biostimulates helping plant develop high plant antioxidant Activity for multiple benefits. *Virginia Vegetable small Fruit and Specialty Crops*. 2 (6) : 1-3.
36. Pandey, N., Pathak, G. C., & Sharma, C. P. (2006). Zinc is critically required for pollen function and fertilisation in lentil. *Journal of Trace Elements in Medicine and Biology*, 20(2), 89-96.
37. Pawłowska K., Janda, K., & Jakubczyk, K. (2020). Properties and use of rosemary (L.). *Pomeranian Journal of Life Sciences*, 66(3), 76-82.
38. Pise, N.M. and A.B. Sabale, 2010. Effect of seaweed concentrates on the growth and biochemical constituents of *Trigonella Foenum-Graecum* L. *J. Phytology*, 2: 50–56.
39. Potter G.(2005) [www. Kaizenbonsai.com](http://www.Kaizenbonsai.com).
40. Prasad, T. N. V. K. V., Sudhakar, P., Sreenivasulu, Y., Latha, P., Munaswamy, V., Reddy, K. R., ... & Pradeep, T. (2012). Effect of nanoscale zinc oxide particles on the germination, growth and yield of peanut. *Journal of plant nutrition*, 35(6), 905-927.
41. Pratt, P. F., & Chapman, H. D. (1961). Gains and losses of mineral elements in an irrigated soil during a 20-year lysimeter investigation.
42. Rayorath, P., Jithesh, M. N., Farid, A., Khan, W., Palanisamy, R., Hankins, S. D., ... & Prithiviraj, B. (2008). Rapid bioassays to evaluate the plant growth promoting activity of *Ascophyllum nodosum* (L.) Le Jol. using a model plant, *Arabidopsis thaliana* (L.) Heynh. *Journal of applied phycology*, 20, 423-429.
43. Reeta, K.A and A.K.Bhatnager(2011).Effect of aqueous extract of *Sargassum johnstonii* Setchell & Gardner on growth , yield and quality of *Lycopersicon esculentum* Mill. *Agronomy .J.Appl Phycol.* 23:623-633.
44. Reitz, S.R. and J. T. Trumble (1996) . Effect of cytokinin containing Seaweed extract on *Phaseolus vulgaris* L. influence of nutrient availability and apex . removal . *Botanica Marine* Vol . 39: 33-38 .
45. Sadeghi, S.; A. Rahnavard and Z. Y. Ashrafi (2009). The effect of plant-density and sowing-date on yield of Basil (*Ocimum basilicum* L.) In Iran. *Journal of Agricultural Technology*, 5(2): 413-422.
46. Saif Eldeen, U. M., & Shokr, M. M. B. (2014). Effect Of Foliar Spray With Seaweeds Extract And Chitozan On Earliness And Productivity Of Globe Artichoke. *Journal of Plant Production*, 5(7), 1197-1207.
47. Seyedbagheri, S. R., Azad, M. M. and Naseri, R. (2021). Influence of foliar application of nano- and chemical manganese sulfate and zinc sulfate on the growth, nutrients, and pigments of green bean (*Phaseolus vulgaris* L.). *Environmental Science and Pollution Research*, 28(9), 10735-10744
48. Shushupti, O., Orpa, R. S., Tarannum, T., Chitra, N. N., Suchi, S. J. H., & Rahman, M. K. (2021). Influence of various commercially available organic manures on growth, yield and nutrient accumulation in mint plants (*Mentha* sp.). *Journal of Biodiversity Conservation and Bioresource Management*, 7(1), 73-84.
49. Soltani, M. (2017). Effect of foliar application of nano-zinc oxide and zinc sulfate on rosemary (*Rosmarinus officinalis* L.) growth and nutrient uptake. *Journal of Plant Nutrition*, 40(11), 1637-1646.
50. Steel RGD , Torrie JH (1980).Principles and procedures of statistics :A Biometrical Approach New York , NY :Mc Graw Hill , USA .
51. Teye, E., Asare, A. P., Amoah, R. S., & Tetteh, J. P. (2011). Determination of the dry matter content of cassava (*Manihot esculenta*, Crantz) tubers using specific gravity method. *ARPN Journal of Agricultural and Biological Science*, 6(11), 23-28

52. Varamin, J. K., Fanoodi, F., Sinaki, J. M., Rezvan, S., & Damavandi, A. (2020). Foliar application of chitosan and nano-magnesium fertilizers influence on seed yield, oil content, photosynthetic pigments, antioxidant enzyme activities of sesame (*Sesamum indicum* L.) under water-limited conditions. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 48(4), 2228-2243.
53. Vernieri, P., Borghesi, E., Ferrante, A., Magnani, G. 2005. Application of biostimulants in floating system for improving rocket quality. *J. Food, Agri. and Environ.* 3(3&4):86- 88.
54. Zhang, X., Ervin, E. H., & Schmidt, R. E. (2003). Physiological effects of liquid applications of a seaweed extract and a humic acid on creeping bentgrass. *Journal of the American Society for Horticultural Science*, 128(4), 492-496.